

# Programming Molecules

Luca Cardelli  
Microsoft Research

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<http://lucacardelli.name>

# Outline

- **Part I: Analyzing molecular networks**
  - We try to discover the function of the network.
  - We try to understand how the structure is dictated by the function (and other natural constraints).
- **Part II: Engineering molecular networks**
  - We know the function we want to implement.
  - We use the structures we have available to implement the function. But we want to do this *in general* (programmatically).

# Part I

## Systems Biology

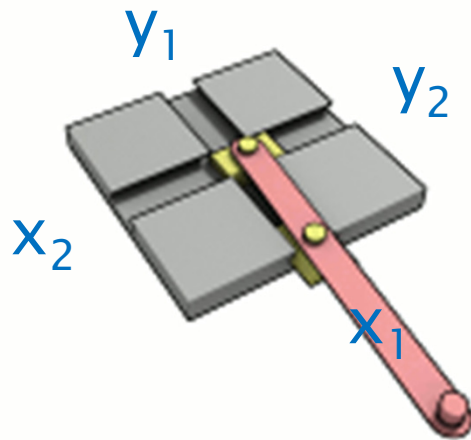
- *or* -

**How Does Nature Build  
Molecular Oscillators?**

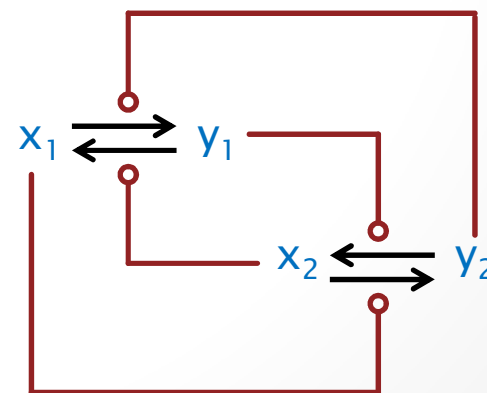
# The Trammel of Archimedes

- A device to draw ellipses
  - Two interconnected switches.
  - When one switch is on (off) it flips the other switch on (off).  
When the other switch is on (off) it flips the first switch off (on).
  - The amplitude is kept constant by mechanical constraints.

The function

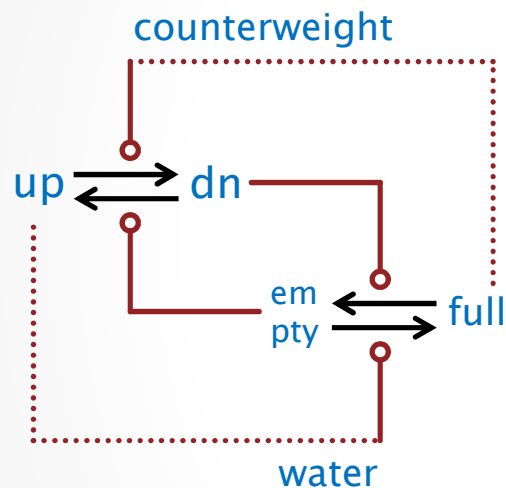


The network

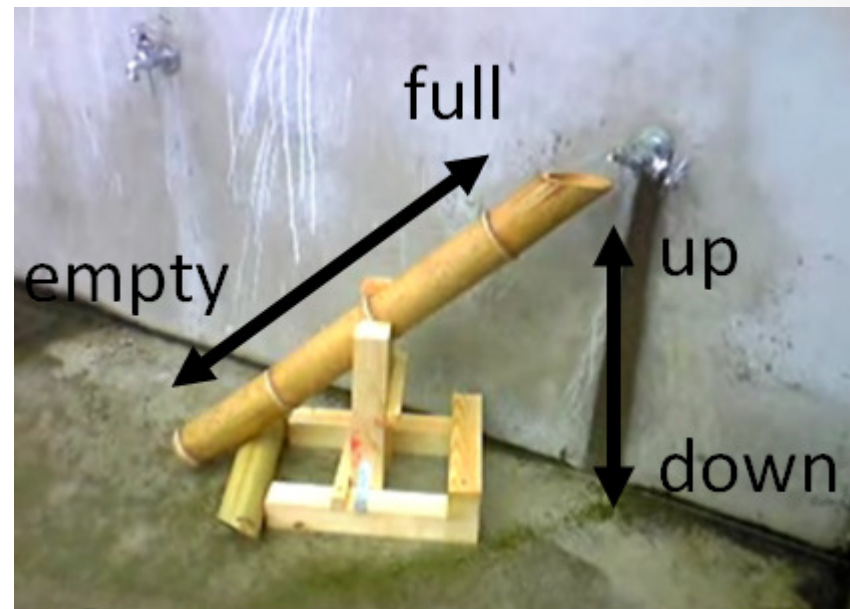


# The Shishi Odoshi

- A Japanese scarecrow (lit. scare-deer)
  - Used by Bela Novak to illustrate the cell cycle switch.



empty + up  $\rightarrow$  up + full  
up + full  $\rightarrow$  full + dn  
full + dn  $\rightarrow$  dn + empty  
dn + empty  $\rightarrow$  empty + up

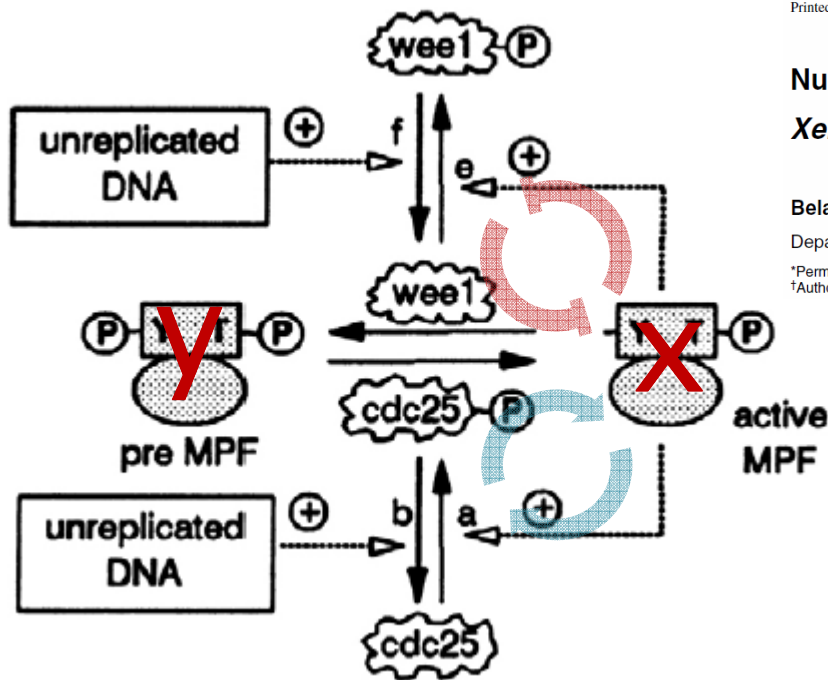


<http://www.youtube.com/watch?v=VbvecTlftcE&NR=1&feature=fwp>

Outer switched connections replaced by constant influxes: tap water and gravity.

# The Cell Cycle Switch

- At the core of the cell-cycled oscillator.
  - This network is universal in all Eukaryotes [P. Nurse].



Journal of Cell Science 106, 1153-1168 (1993)  
Printed in Great Britain © The Company of Biologists Limited 1993

Numerical analysis of a comprehensive model of M-phase control in *Xenopus* oocyte extracts and intact embryos

Bela Novak\* and John J. Tyson†

Department of Biology, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24060-0406, USA

\*Permanent address: Department of Agricultural Chemical Technology, Technical University of Budapest, 1521 Budapest Gellert Ter 4, Hungary  
†Author for correspondence

- Double positive feedback on x
- Double negative feedback on x
- No feedback on y
- ???

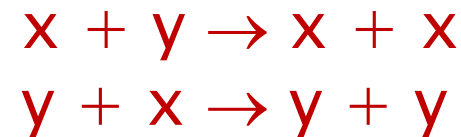
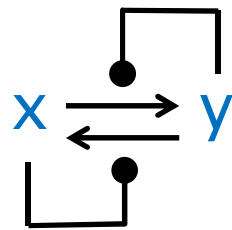
- Well studied. But why this structure?

# How to Build a Switch

- What is a “good” switch?
  - We need first a *bistable* system: one that has two *distinct* and *stable* states. I.e., given *any* initial state the system must *settle* into one of two states.
  - The settling must be *fast* (not get stuck in the middle for too long) and *robust* (must not spontaneously switch back).
  - Finally, we need to be able to *flip* the switch: drive the transitions by external inputs.

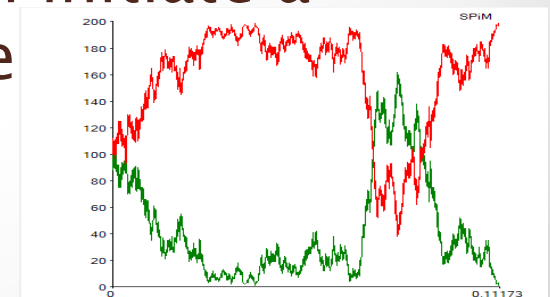
# A Bad Algorithm

- Direct x-y competition
  - x catalyzes the transformation of y into x
  - y catalyzes the transformation of x into y



- This system is bistable, but
  - Convergence to a stable state is *slow* (a random wa
  - *Any* perturbation of a stable state can initiate a random walk to the other stable state

```
objective sample 0.0002
1000
direction plot x(t), y(t)
end of = 1.0
new sp: update: time
new sp: update: time
set x() =
set y() = 0
or 'set' x()
end of =
do 'set' x()
or 'set' x()
end of =
do 'set' x()
or 'set' x()
run 100 of x()
run 100 of y()
```





# A Very Good Algorithm

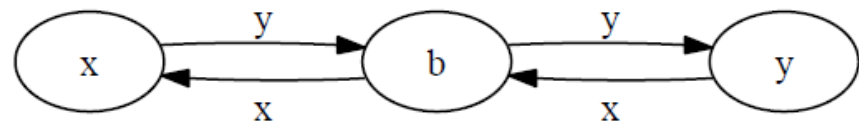
- Approximate Majority
  - Decide which of two populations is in majority
- A fundamental ‘population protocol’
  - Agents in a population start in state  $x$  or state  $y$ .
  - A pair of agents is chosen randomly at each step, they interact ("collide") and change state.
  - The whole population must eventually agree on a majority value (all  $x$  or all  $y$ ) with probability 1.

Dana Angluin · James Aspnes · David Eisenstat

## A Simple Population Protocol for Fast Robust Approximate Majority

We analyze the behavior of the following population protocol with states  $Q = \{b, x, y\}$ . The state  $b$  is the **blank** state. Row labels give the initiator's state and column labels the responder's state.

	$x$	$b$	$y$
$x$	$(x, x)$	$(x, x)$	$(x, b)$
$b$	$(b, x)$	$(b, b)$	$(b, y)$
$y$	$(y, b)$	$(y, y)$	$(y, y)$



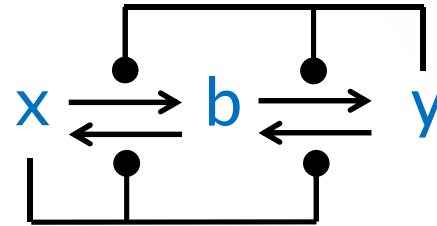
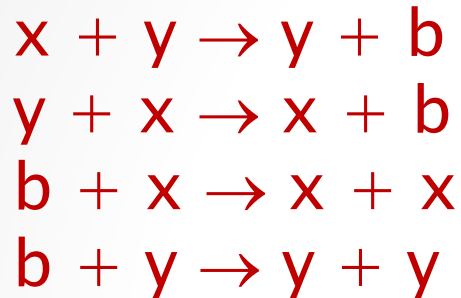
Third ‘undecided’ state.

# Properties

- With high probability, for  $n$  agents [Angluin et al. <http://www.cs.yale.edu/homes/aspnes/papers/disc2007-eisenstat-slides.pdf>]
  - The number of state changes before converging is  $O(n \log n)$
  - The total number of interactions before converging is  $O(n \log n)$
  - The final outcome is correct if the initial disparity is  $\omega(\sqrt{n \log n})$
- The algorithm is the fastest possible
  - Must wait  $\Omega(n \log n)$  steps in expectation for all agents to interact
- Logarithmic time bound
  - Parallel time is the number of steps divided by the number of agents.
  - In parallel time the algorithm converges with high probability in  $O(\log n)$ .
  - That is true for any initial conditions, even  $x=y!$

“Although we have described the population protocol model in a sequential light, in which each step is a single pairwise interaction, interactions between pairs involving different agents are independent and may be thought of as occurring in parallel. In measuring the speed of population protocols, then, we define 1 unit of parallel time to be  $\sum_j j$  steps. The rationale is that in expectation, each agent initiates 1 interaction per parallel time unit; this corresponds to the chemists’ idealized assumption of a well-mixed solution.”  
Distributed Computing 21(2):87-102.

# Chemical Implementation



Worse case test: start with  $x=y$ .

Bistable

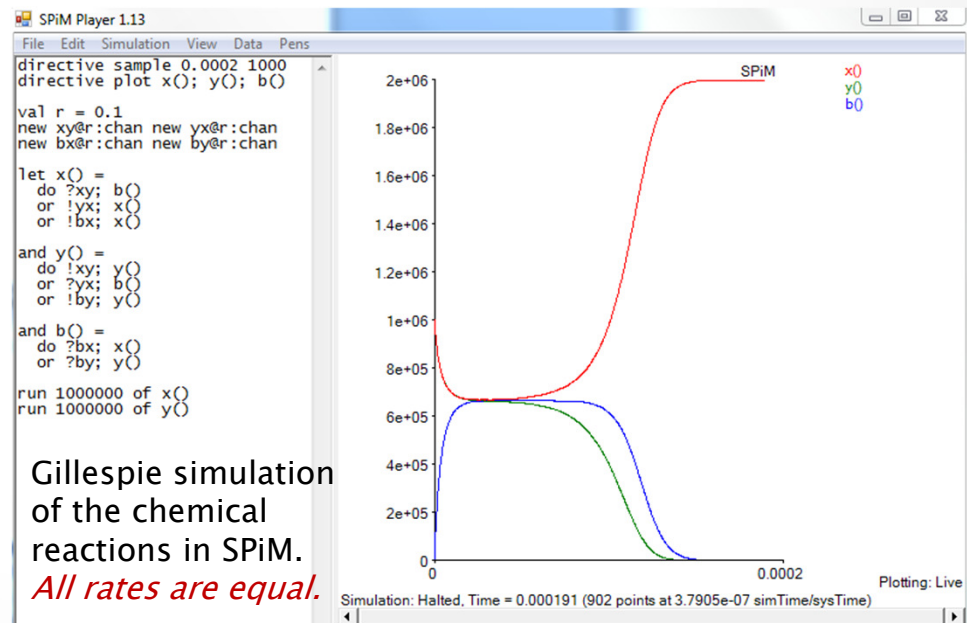
Even when  $x=y$ ! (stochastically)

Fast

$O(\log n)$  convergence time

Robust

$\omega(\sqrt{n} \log n)$  majority wins whp

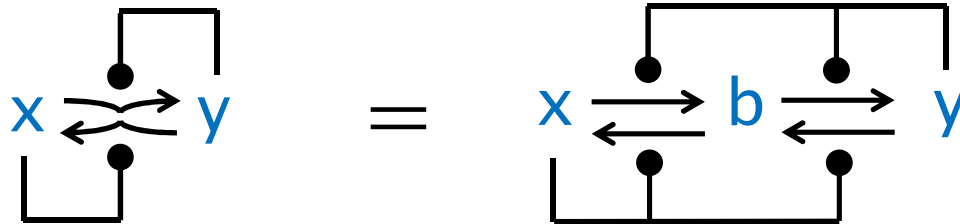


# Back to the Cell Cycle

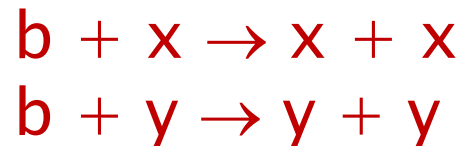
- The AM algorithm has great properties for settling a population into one of two states.
- But that is not what the cell cycle uses to switch its populations of molecules.
- Or is it?

# Step 1: the AM Network

*Abbreviated notation:*

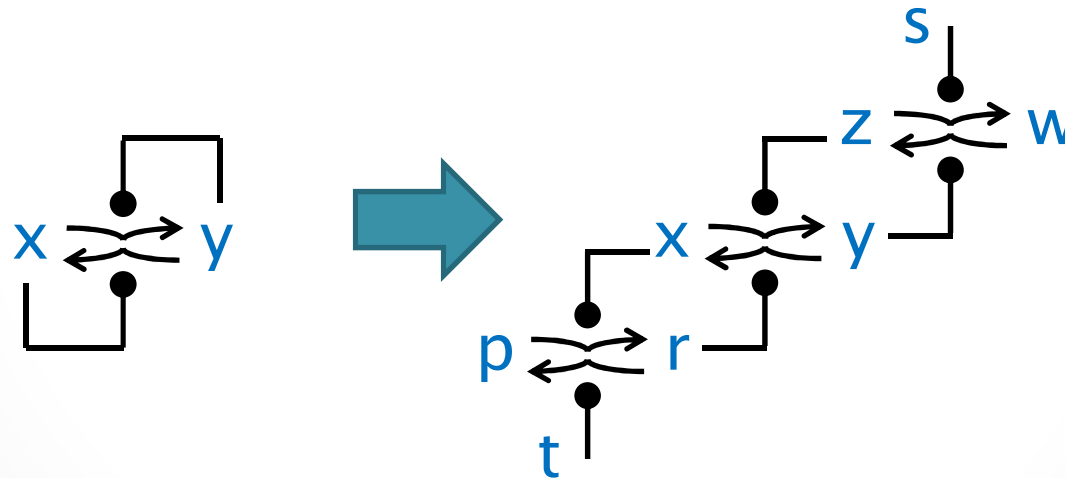


- Autocatalysis, and especially intricate autocatalysis, is not commonly seen in nature. Presumably, it's hard:



# Step 2: remove auto-catalysis

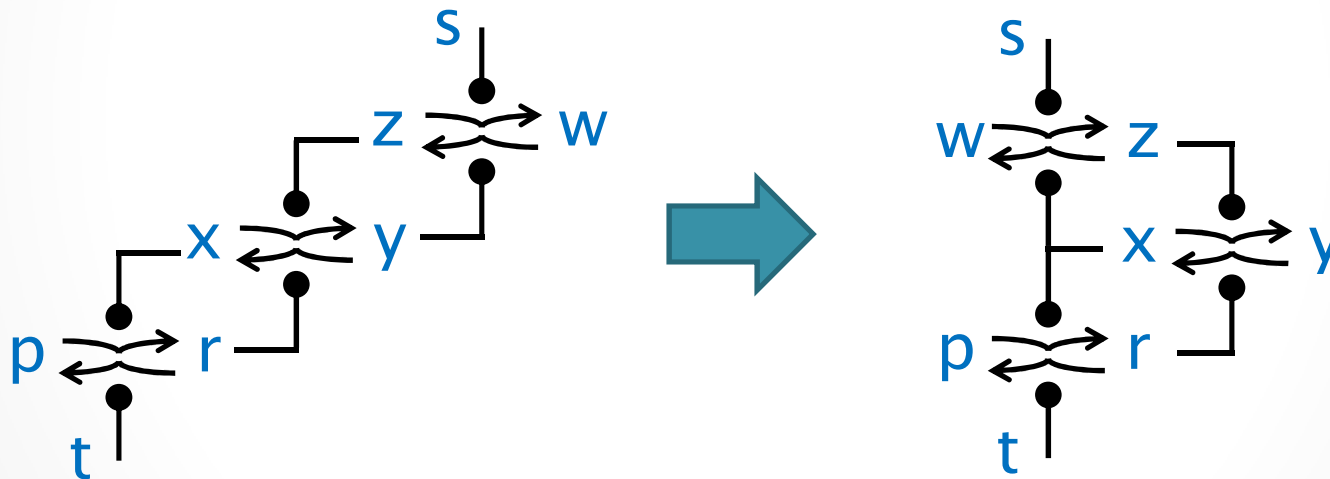
- Replace autocatalysis by mutual (simple) catalysis, introducing intermediate species z, r.
  - Here z breaks the y auto-catalysis, and r breaks the x auto-catalysis, while preserving the feedbacks.
  - z and r need to 'relax back' (to w and p) when they are not catalyzed: s and t provide the back pressure.



- Still, x and y (two states of the same molecule) are distinct active catalysts: that is not common in nature either.

# Step 3: only one active state

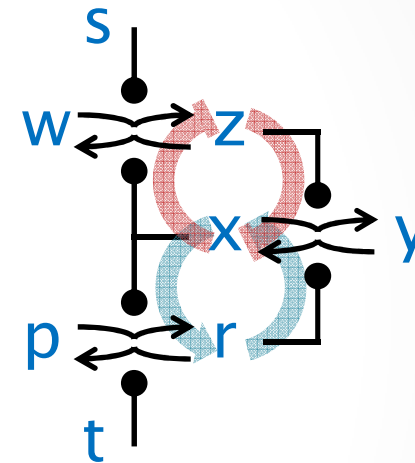
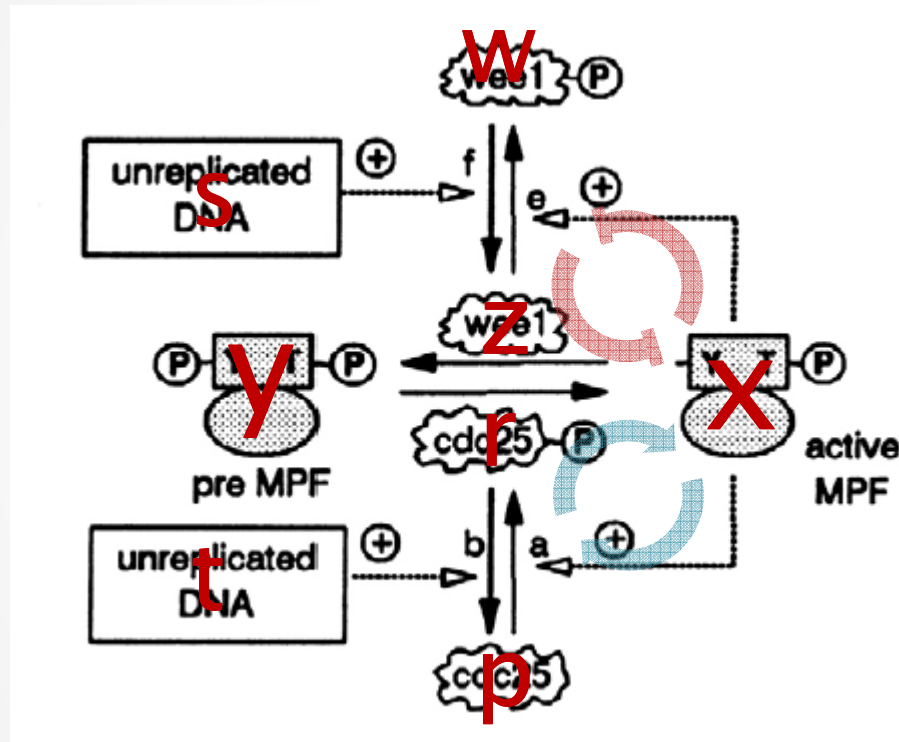
- Remove the catalytic activity of  $y$ .
  - Instead of  $y$  activating itself through  $z$ , we are left with  $z$  activating  $y$  (which remains passive). Hence, to deactivate  $y$  we now need to deactivate  $z$ . Since  $x$  'wants' to deactivate  $y$ , we make  $x$  deactivate  $z$ .



- All species now have one active ( $x, z, r$ ) and one inactive ( $y, w, p$ ) form. This is 'normal'.

# Network Structure

- ... and that *is* the cell-cycle switch!



(Some of the bistable states can be enzymatic rather than multi-site phosphorylations as in AM.)

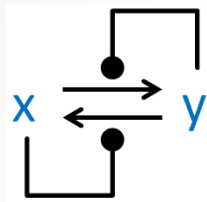
- The question is: did we preserve enough *function* through our *network transformations*?



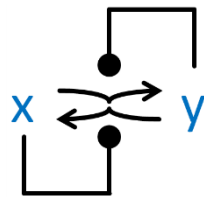
# Quantitative Analysis

Switches as Computational Systems – Convergence

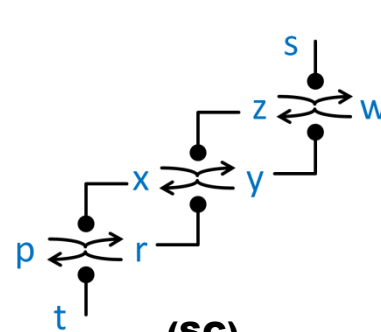
Techniques: Stochastic Simulation and Probabilistic Modelchecking



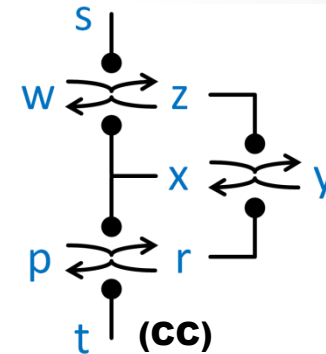
(DC)



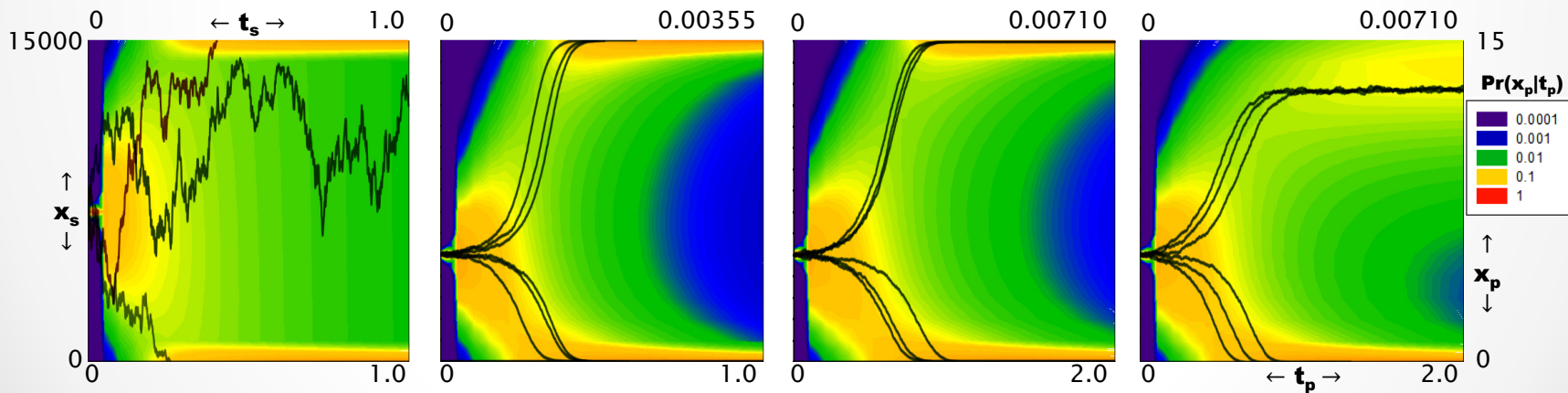
(AM)



(SC)



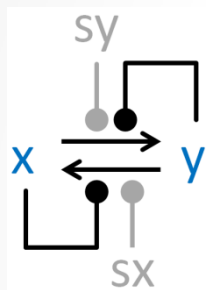
(CC)



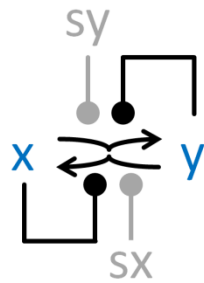
Joint work with Attila Csikász-Nagy

# Quantitative Analysis

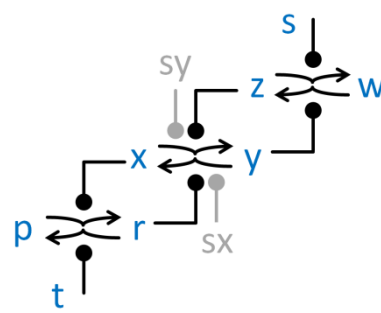
Switches as Dynamical Systems – Steady State Response  
**Techniques:** as above, plus Dynamical Systems Theory



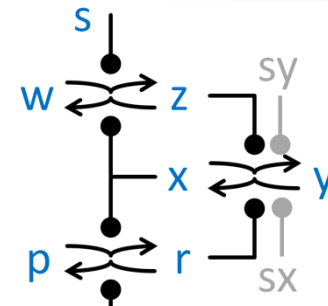
(DC)



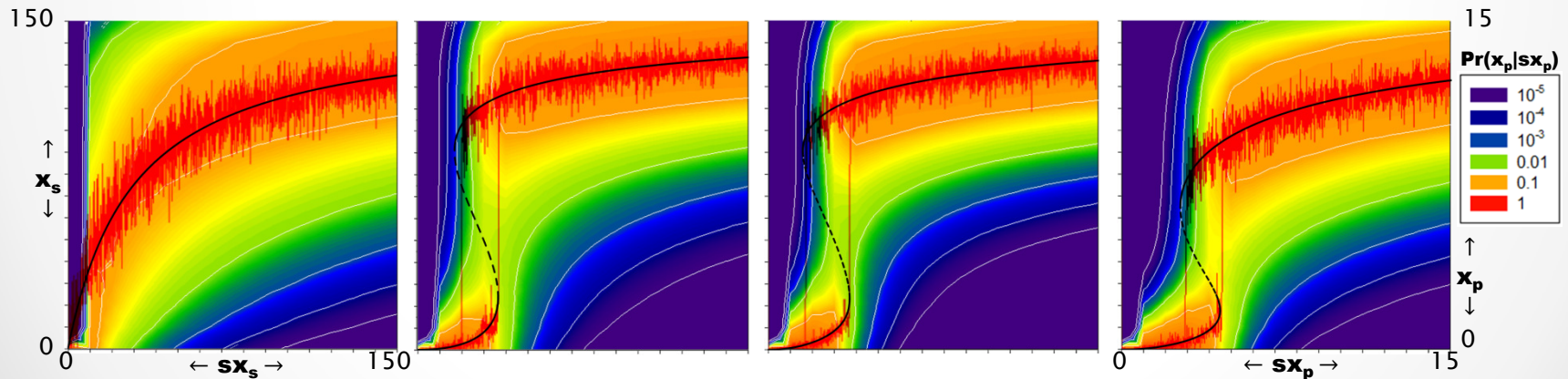
(AM)



(SC)



(CC)



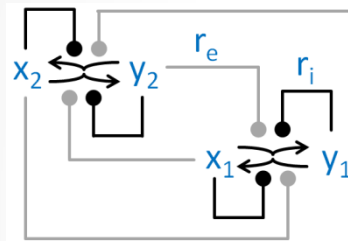
Joint work with Attila Csikász-Nagy

# Quantitative Analysis

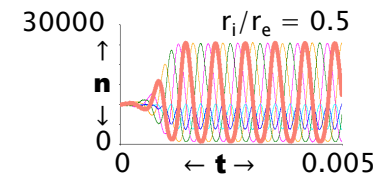
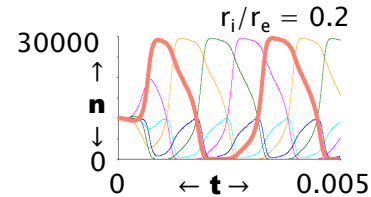
Switches in the context of larger networks

**Techniques: testing**

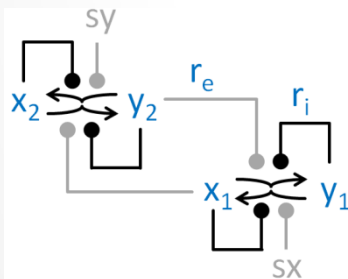
(We have better techniques for non-quantitative systems.)



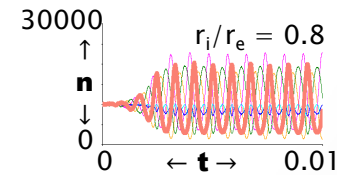
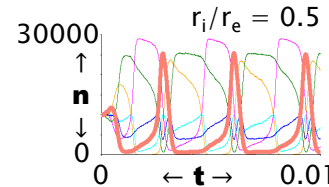
**Trammel**



x1  
y1  
b1  
x2  
y2  
b2



**Shishi Odoshi**



x1  
y1  
b1  
x2  
y2  
b2

Joint work with Attila Csikász-Nagy

# Summary

- Q (traditional): What kind of dynamical system is the cell-cycle switch?
- A (traditional): Bistability – ultrasensitivity – hysteresis ...  
Focused on how unstructured sub-populations change over time.
  
- Q: What kind of algorithmic system is the cell-cycle switch?
- A: Interaction – complexity – convergence ...  
Focused on individual molecules as programmable, structured, algorithmic entities.

# Part II

## Synthetic Biology

- *or* -

How Can We Build  
Molecular Oscillators?  
(or any other network?)

# Molecular Programming Languages

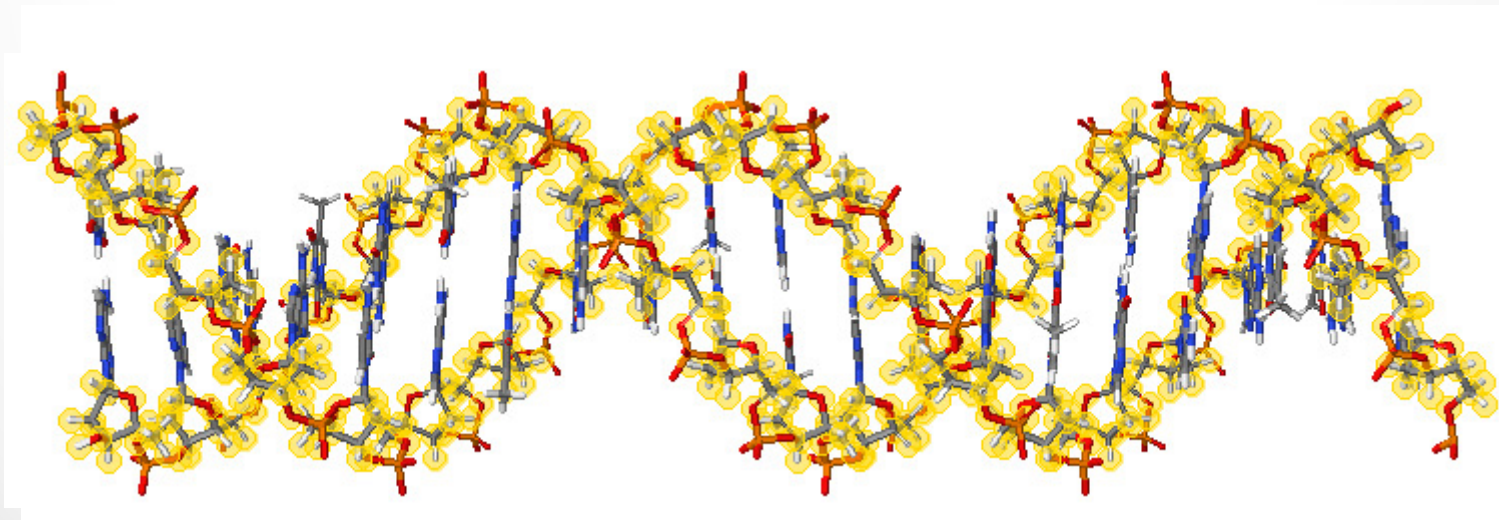
- Reaction-Based ( $A + B \rightarrow C + D$ ) (Chemistry)
  - Limited to finite set of species (no polymerization)
  - Practically limited to small number of species (no run-away complexation)
- Interaction-Based ( $A = !r; C$ ) (Process Algebra)
  - Reduces combinatorial complexity of models by combining independent submodels connected by interactions.
- Rule-Based ( $A\{-\}:B\{p\} \rightarrow A\{p\}:B\{-\}$ ) (Logic, Graph Rewriting)
  - Further reduces model complexity by describing molecular state, and by allowing one to ‘ignore the context’: a *rule* is a reaction in an unspecified (complexation/phosphorylation) context.
  - Similar to informal descriptions of biochemical events (“narratives”).
- Different levels of representation efficiency
  - The latter two can be translated (to each other and) to the first, but doing so may introduce an infinite, or anyway *extremely large*, number of species.

# But what about Execution?

- Chemistry is not easily executable
  - Please Mr Chemist, execute me these reactions that I just made up.
- Description
  - Molecular languages used in systems biology are **descriptive** (modeling) languages
- Compilation
  - How can we **compile** *arbitrary* molecular programs?
- Execution
  - How can we actually **execute** molecular languages? With real molecules?

# DNA as an Engineering Material

- This is why DNA/RNA is important: it is **programmable matter**.
- Not the only one, in principle, but the only one for which we have a well-developed manufacturing technology.



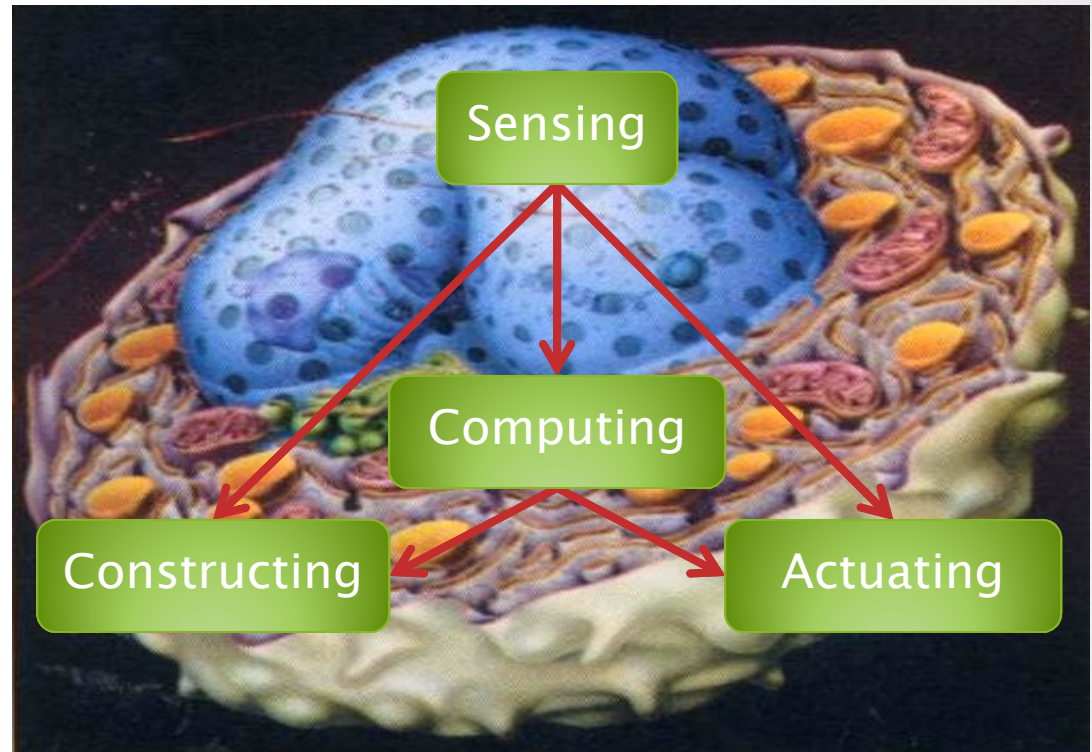
Sequence of Base Pairs (GACT alphabet)



# Molecular Control Systems

- **Sensing**
  - Reacting to forces
  - Binding to molecules
- **Actuating**
  - Releasing molecules
  - Producing forces
- **Constructing**
  - Chassis
  - Growth
- **Computing**
  - Signal Processing
  - Decision Making

## Control Systems



Nucleic Acids can do all this.  
And interface to **biology**.

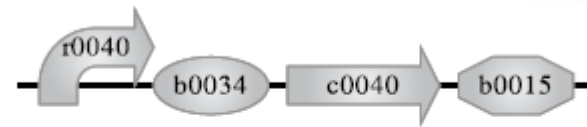
# “Embedded” DNA Computing

(Synthetic Biology)

- Using bacterial machinery (e.g.) as the hardware.  
Using embedded gene networks as the software.
- MIT Registry of Standard Biological Parts

- **GenoCAD**

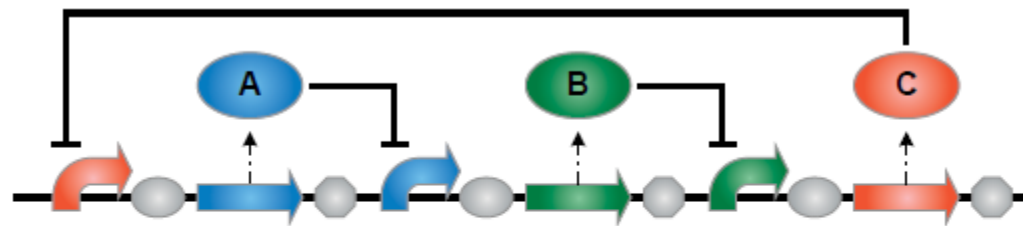
- Meaningful sequences [Cai et al.]



r0040:prom; b0034:rbs; c0040:pcr; b0015:ter

- **GEC**

- [Pedersen & Phillips]



```
prom<neg (C)>; rbs; pcr<codes (A)>; ter;  
prom<neg (A)>; rbs; pcr<codes (B)>; ter;  
prom<neg (B)>; rbs; pcr<codes (C)>; ter
```

# “Autonomous” DNA Computing

(Nano-engineering with biological materials)

- Mix & go
  - All (or most) parts are synthesized
  - No manual cycling (cf. early DNA computing)
  - In some cases, all parts are made of DNA (no enzyme/proteins)
  
- Self-assembled and self-powered
  - Can run on its own (e.g. environmental sensing)
  - Or be embedded into organisms (in the future)

# Curing

A doctor in each cell

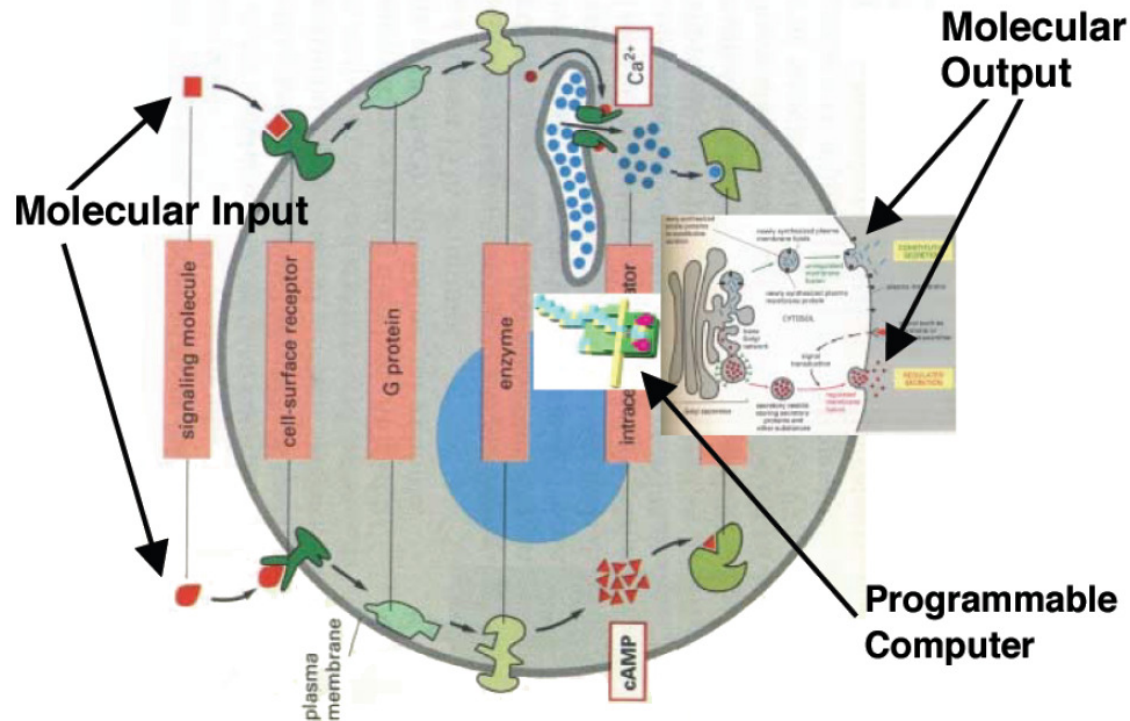
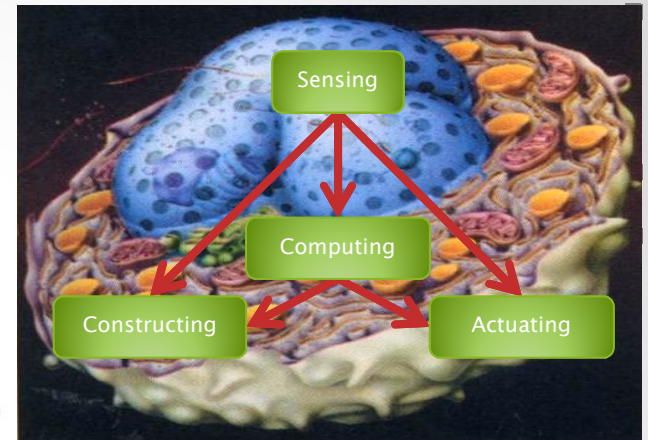


Fig. 1 Medicine in 2050: "Doctor in a Cell"

Ehud Shapiro

Rivka Adar  
Kobi Benenson  
Gregory Linshitz  
Aviv Regev  
William Silverman

**Molecules and  
computation**

# Modern DNA Computing

- Non-goals
  - Not to solve NP-complete problems.
  - Not to replace electronic computers.
  - Not necessarily using genes or to producing proteins.
- For general ‘molecular programming’
  - To precisely control the organization and dynamics of matter and information at the molecular level.
  - To interact algorithmically with biological entities.

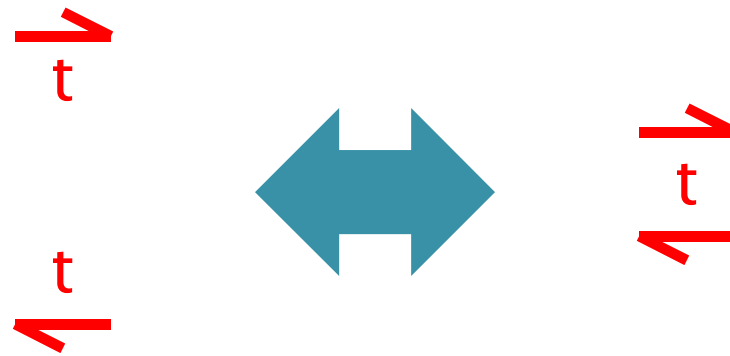
# Domains

- Subsequences on a DNA strand are called **domains**. *PROVIDED* they are “independent” of each other.



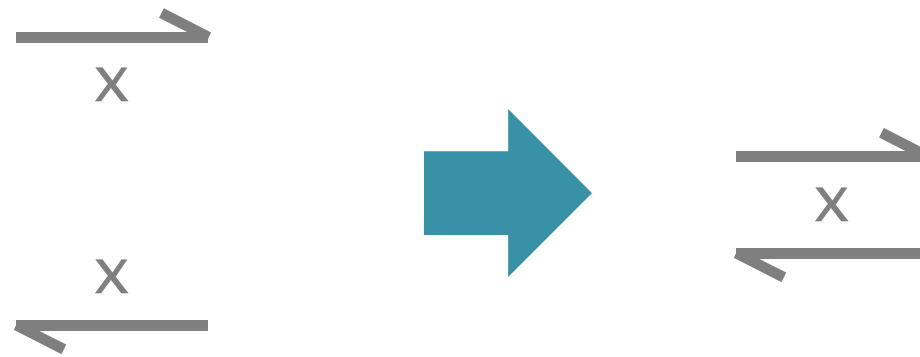
- I.e., differently named domains must not hybridize:
  - With each other
  - With each other's complement
  - With subsequences of each other
  - With concatenations of other domains (or their complements)
  - Etc.
- Choosing domains (subsequences) that are suitably independent is a tricky issue that is still somewhat of an open problem (with a vast literature). But it can work in practice.

# Short Domains



Reversible Hybridization

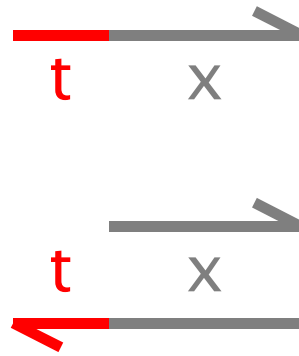
# Long Domains



Irreversible Hybridization

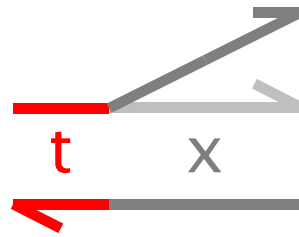


# Strand Displacement



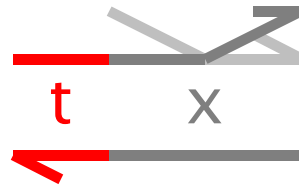
“Toehold Mediated”

# Strand Displacement



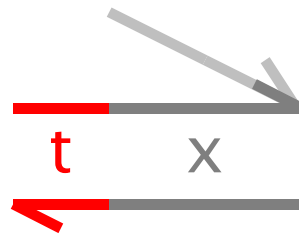
Toehold Binding

# Strand Displacement



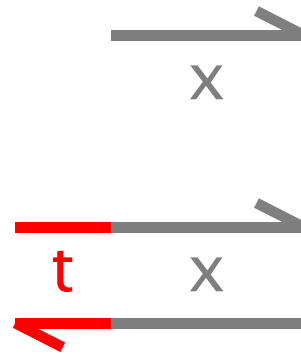
Branch Migration

# Strand Displacement



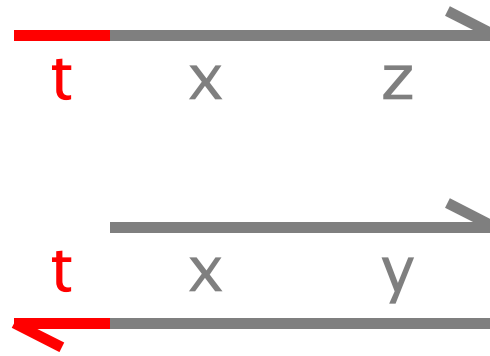
Displacement

# Strand Displacement

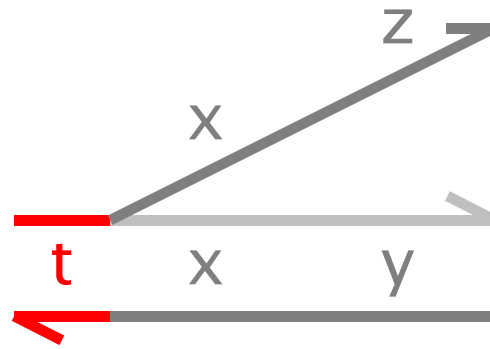


Irreversible release

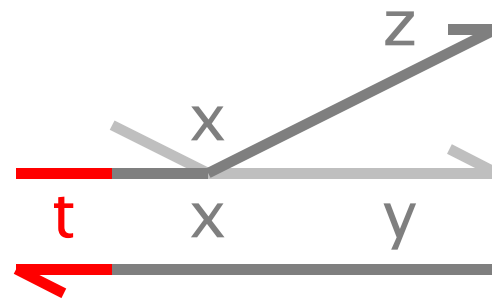
# Bad Match



# Bad Match

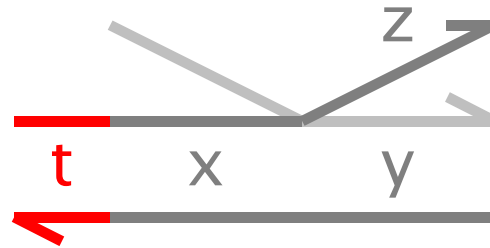


# Bad Match





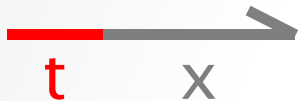
# Bad Match



Cannot proceed  
Hence will undo

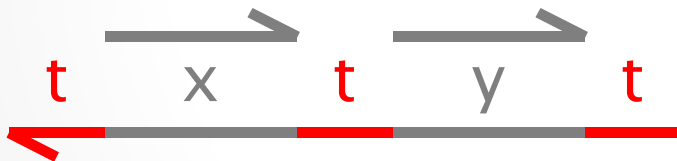
# Two-Domain Architecture

- Signals: 1 toehold + 1 recognition region



Garbage collection  
“built into” the gates

- Gates: “top-nicked double strands”  
(or equivalently double strands with open toeholds)

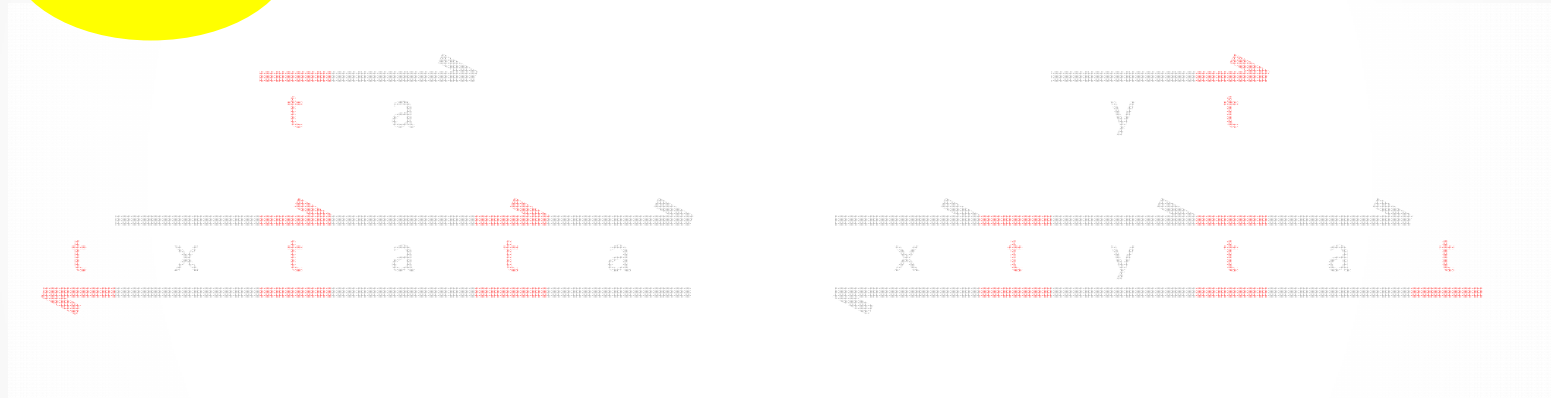
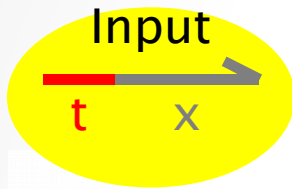


## Two-Domain DNA Strand Displacement

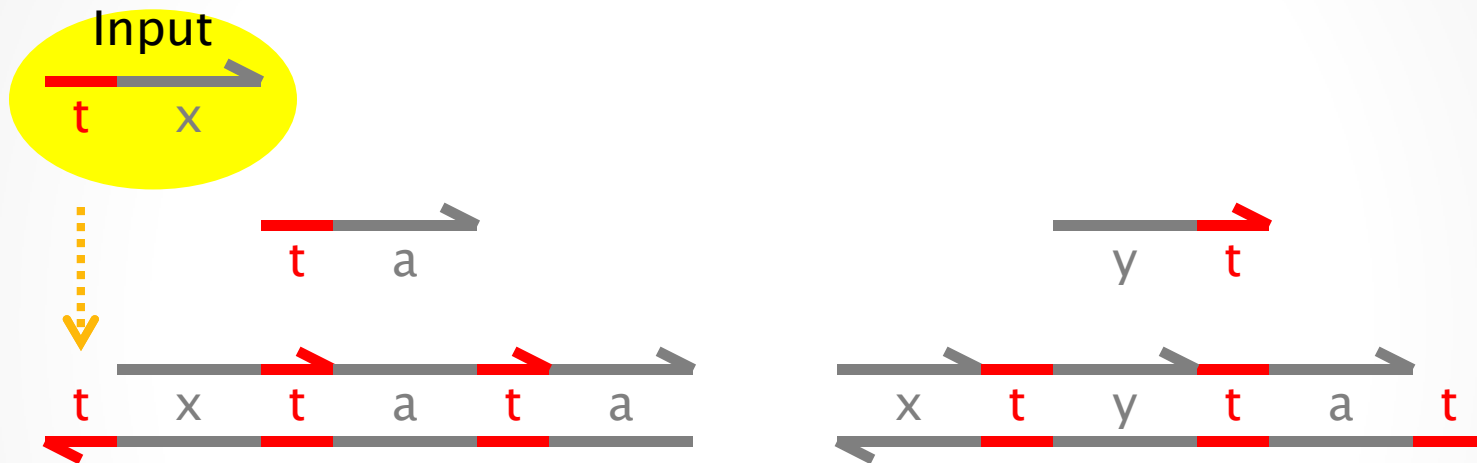
*Luca Cardelli*

In S. B. Cooper, E. Kashefi, P. Panangaden (Eds.):  
Developments in Computational Models (DCM 2010).  
EPTCS 25, 2010, pp. 33–47. May 2010.

# Transducer $x \rightarrow y$



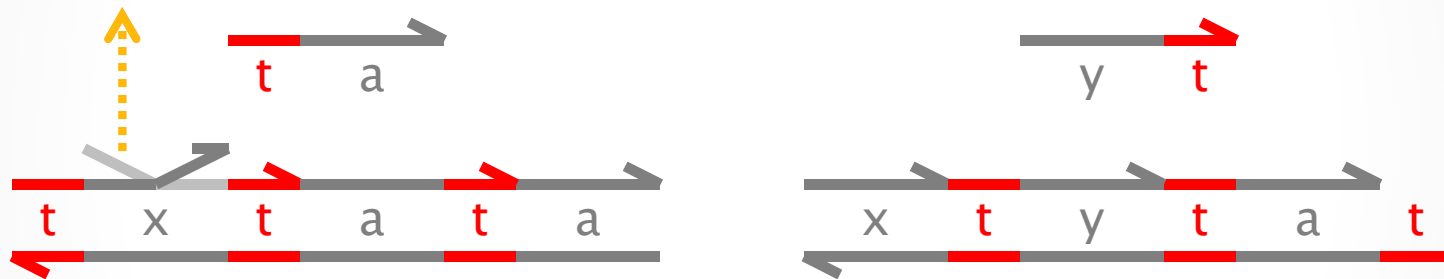
# Transducer $x \rightarrow y$



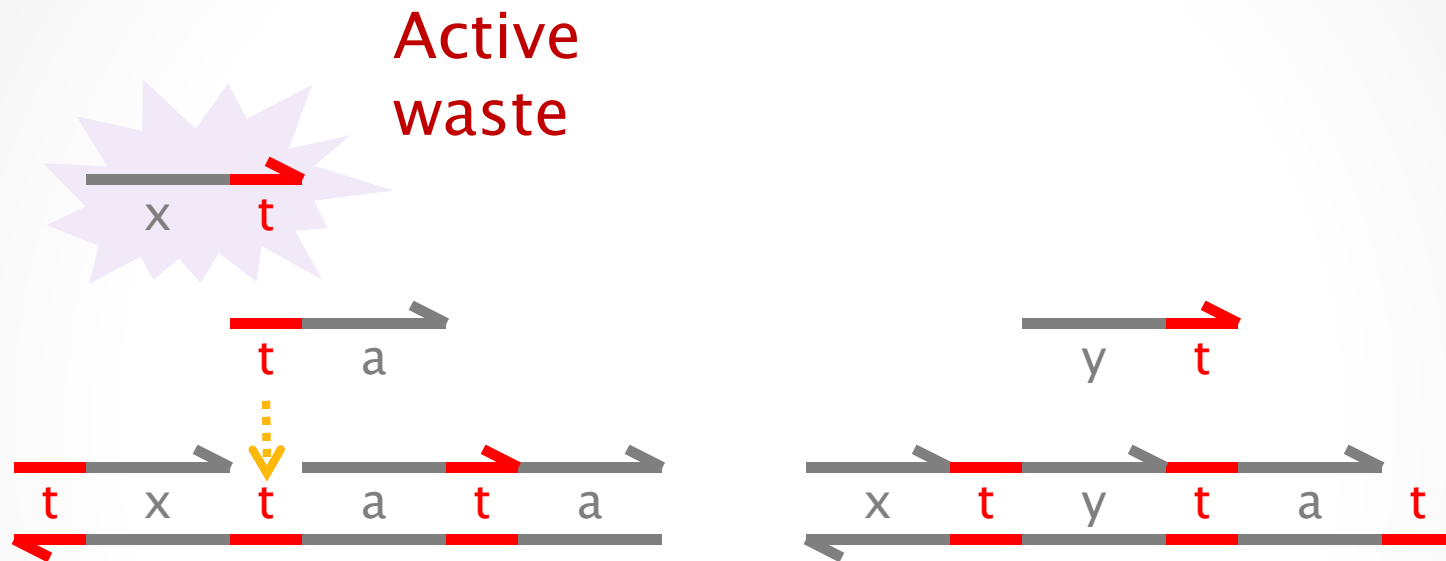
**Built by self-assembly!**

$ta$  is a *private* signal (a different 'a' for each  $xy$  pair)

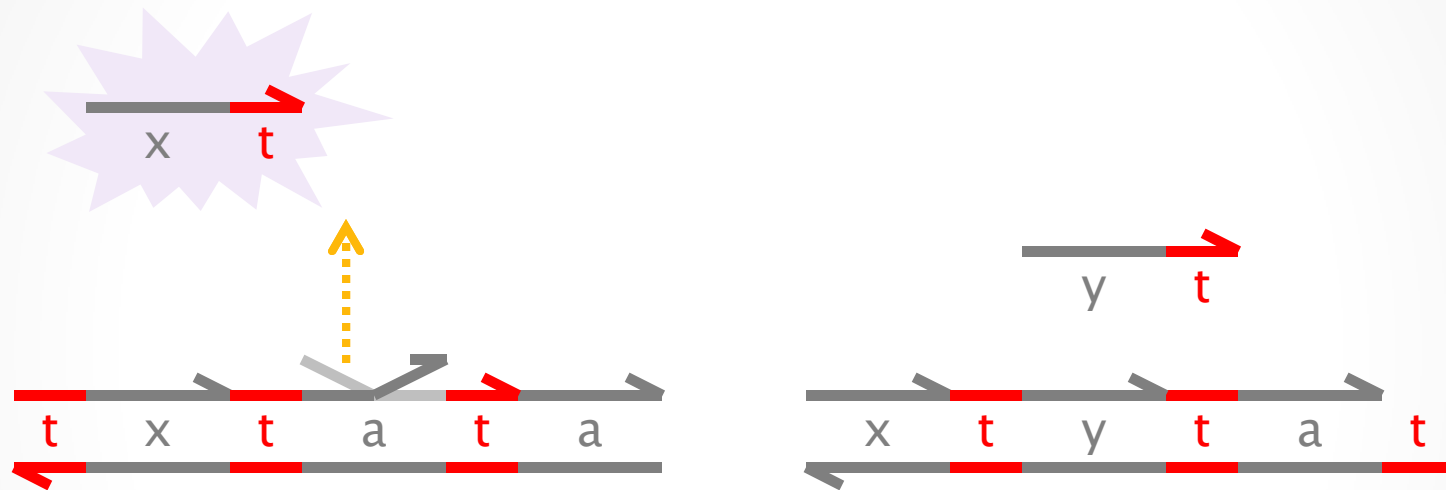
# Transducer $x \rightarrow y$



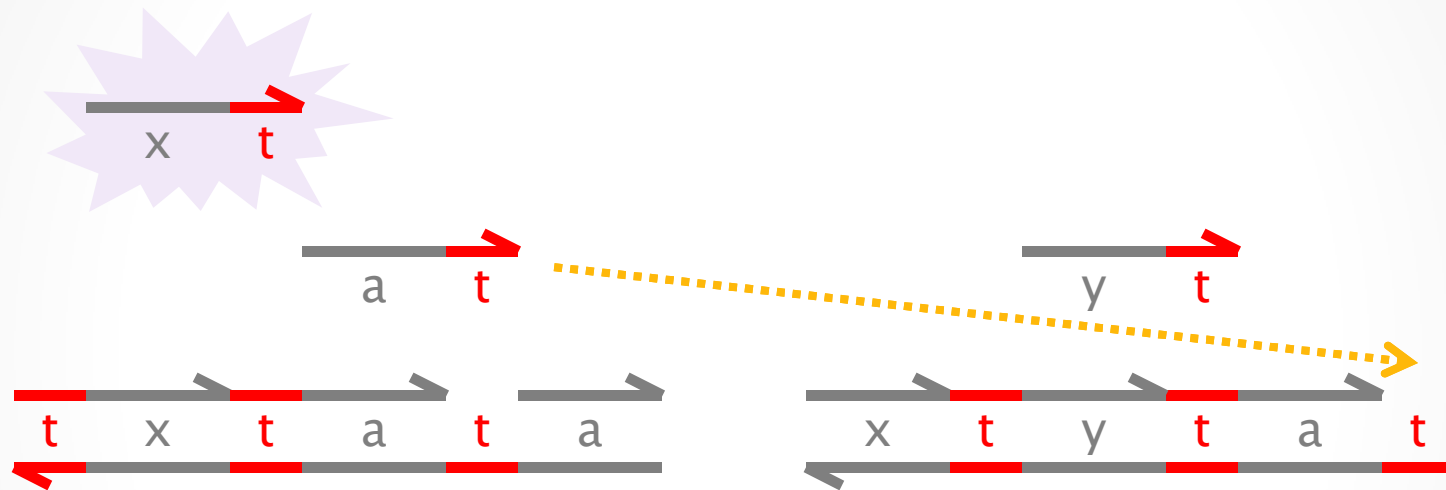
# Transducer $x \rightarrow y$



# Transducer $x \rightarrow y$



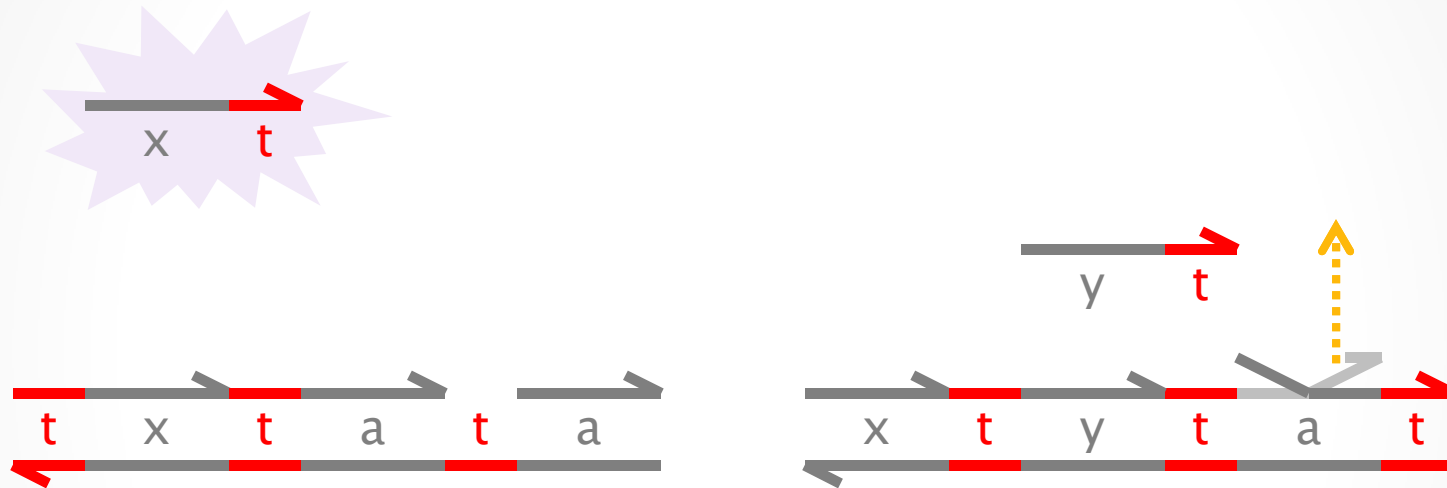
# Transducer $x \rightarrow y$



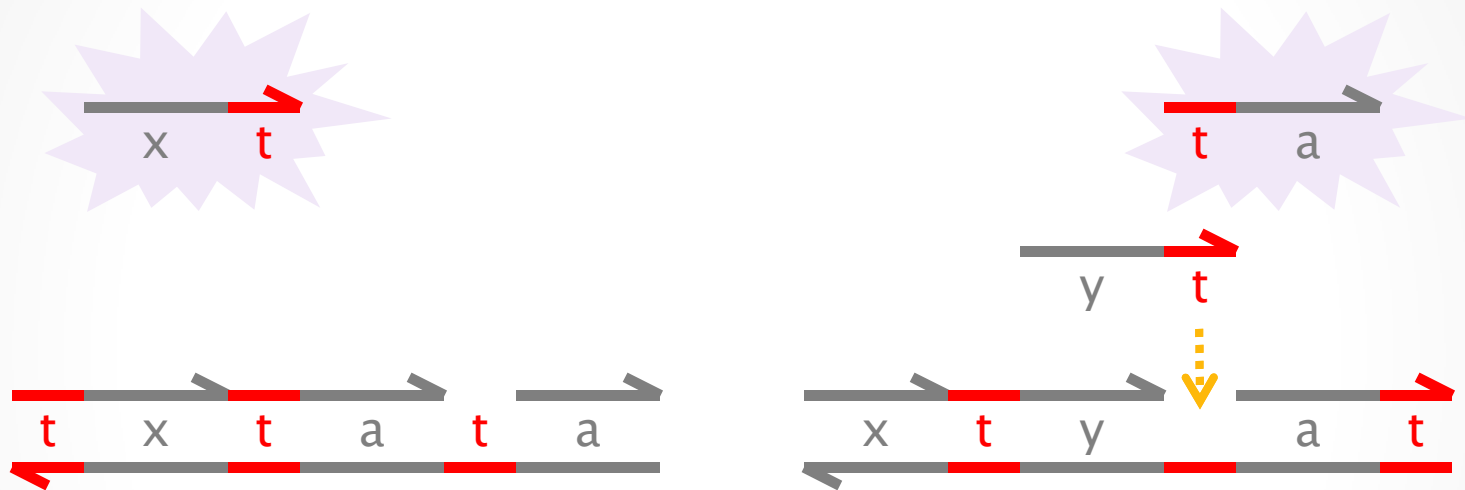
So far, a  $tx$  *signal* has produced an  $at$  *cosignal*.  
But we want signals as output, not cosignals.



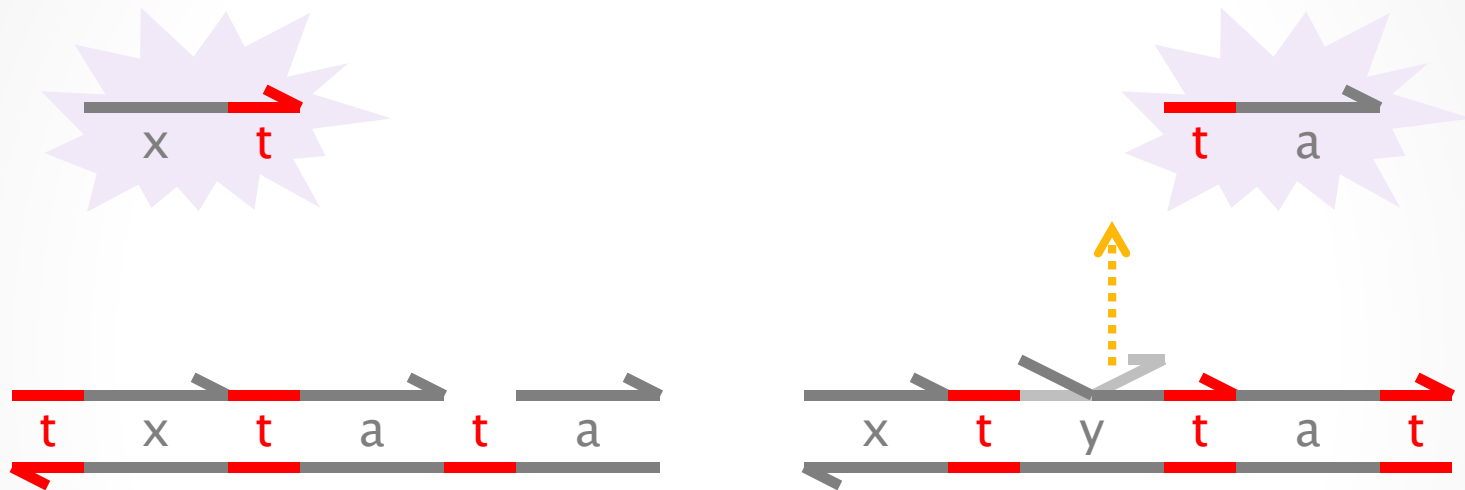
# Transducer $x \rightarrow y$



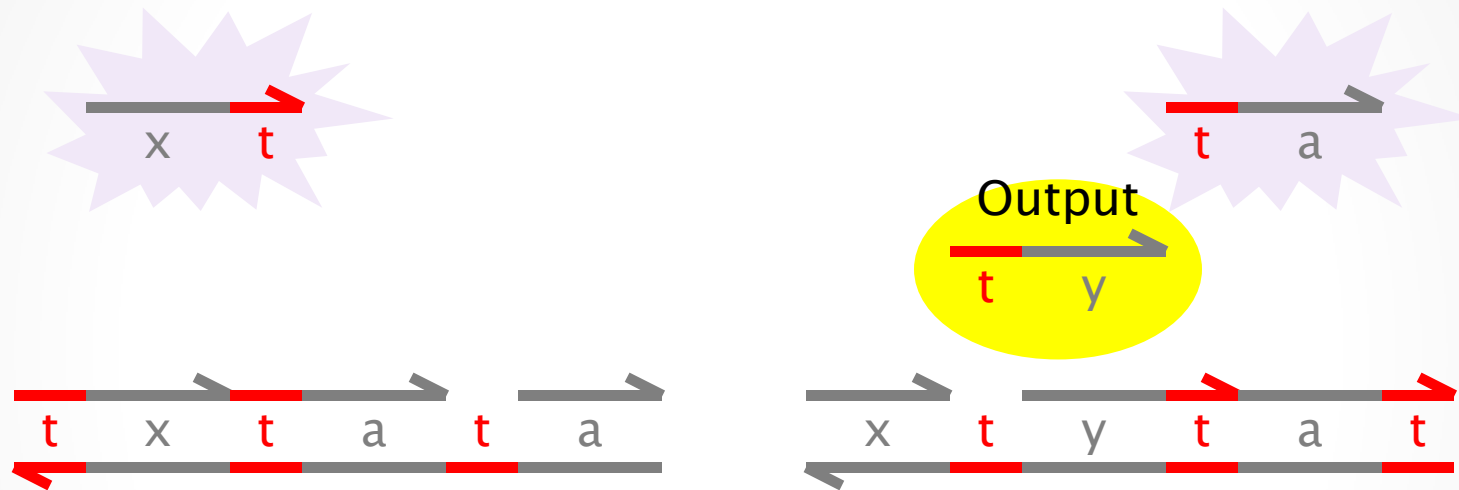
# Transducer $x \rightarrow y$



# Transducer $x \rightarrow y$



# Transducer $x \rightarrow y$



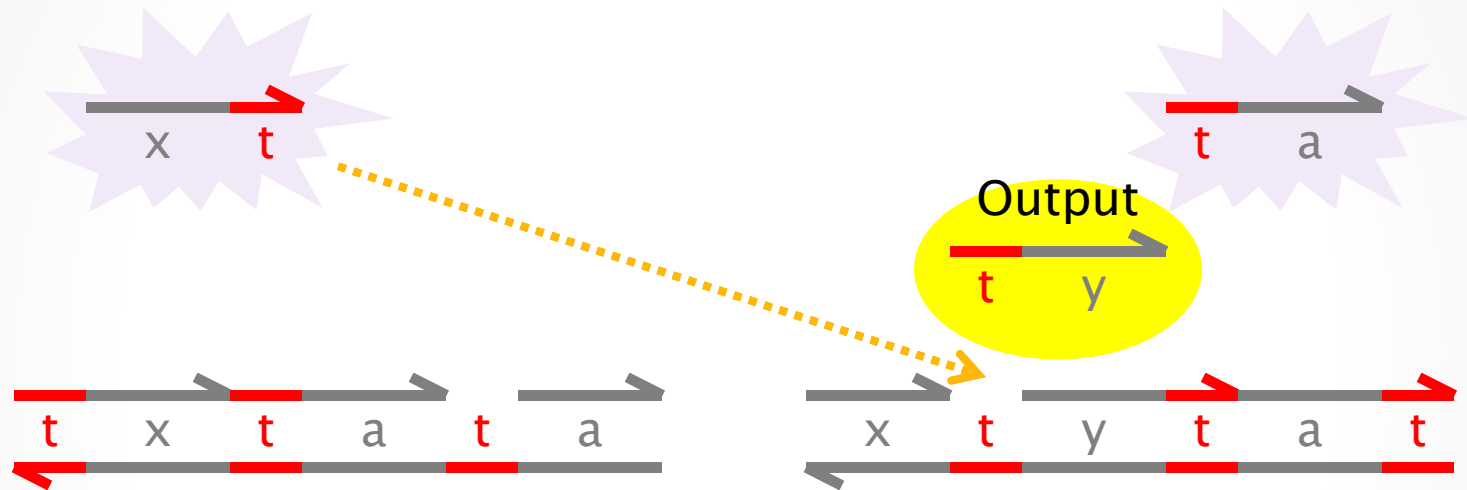
Here is our output *ty signal*.

But we are not done yet:

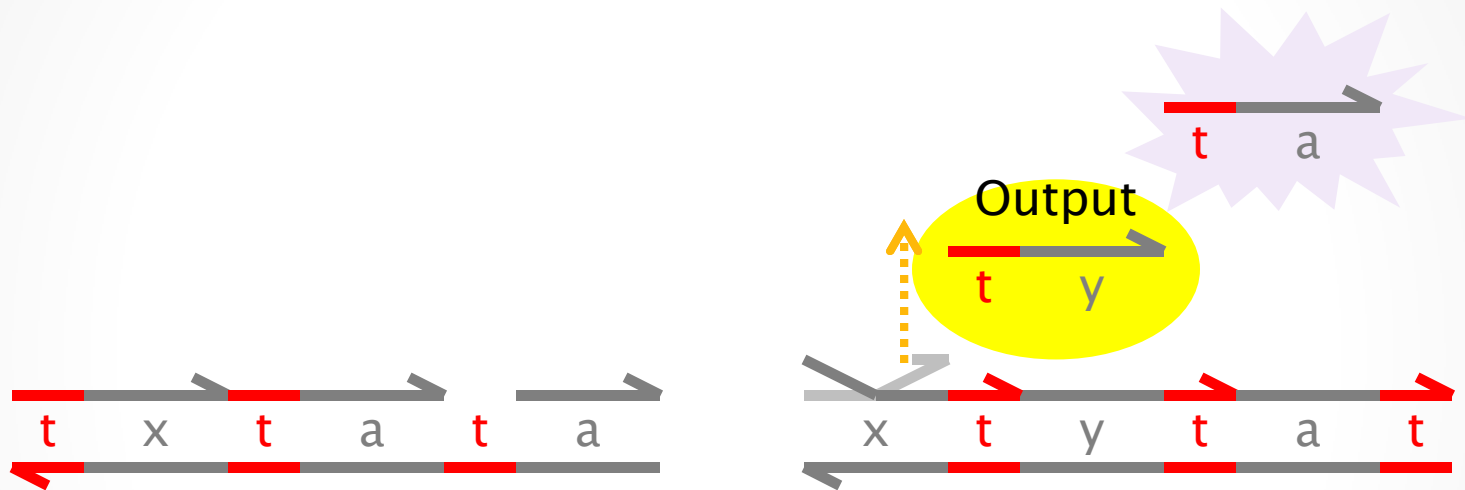
- 1) We need to make the output irreversible.
- 2) We need to remove the garbage.

We can use (2) to achieve (1).

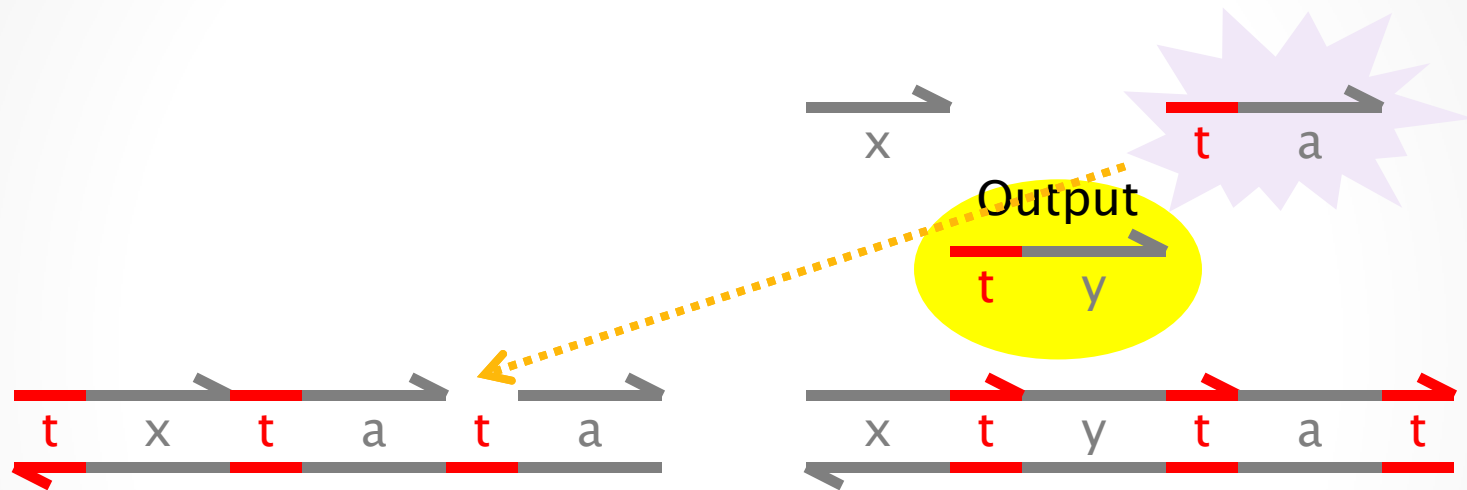
# Transducer $x \rightarrow y$



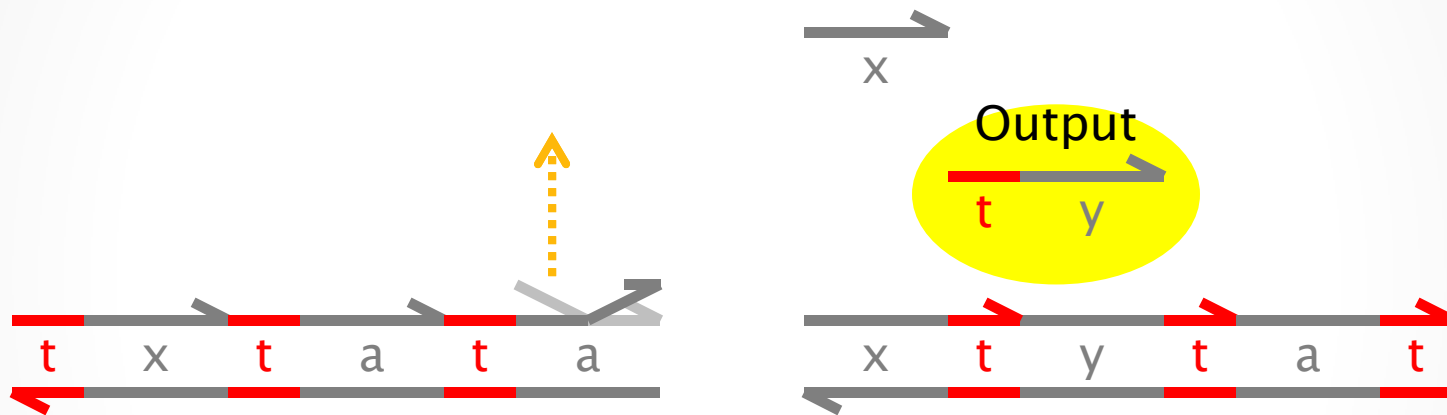
# Transducer $x \rightarrow y$



# Transducer $x \rightarrow y$

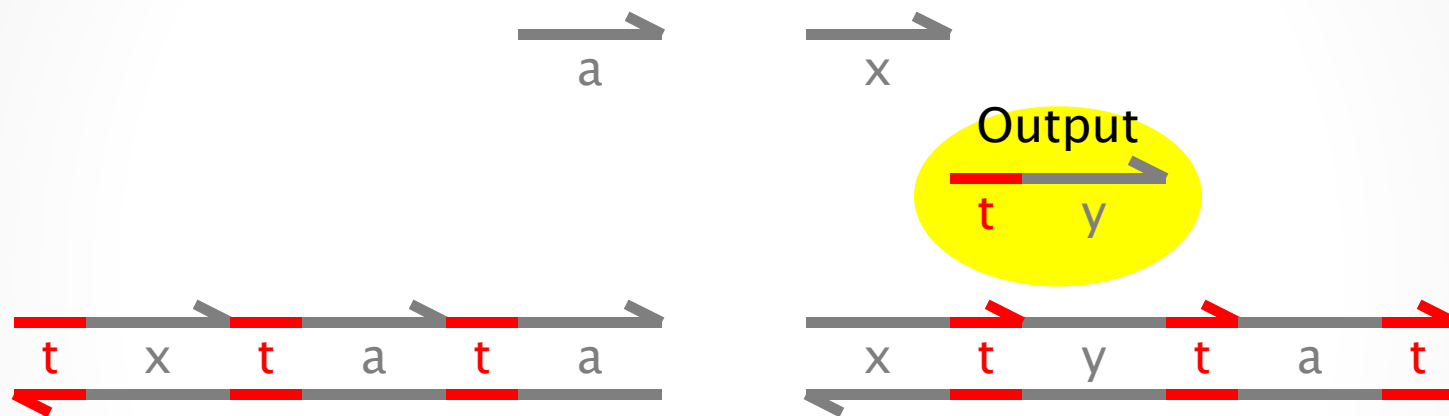


# Transducer $x \rightarrow y$

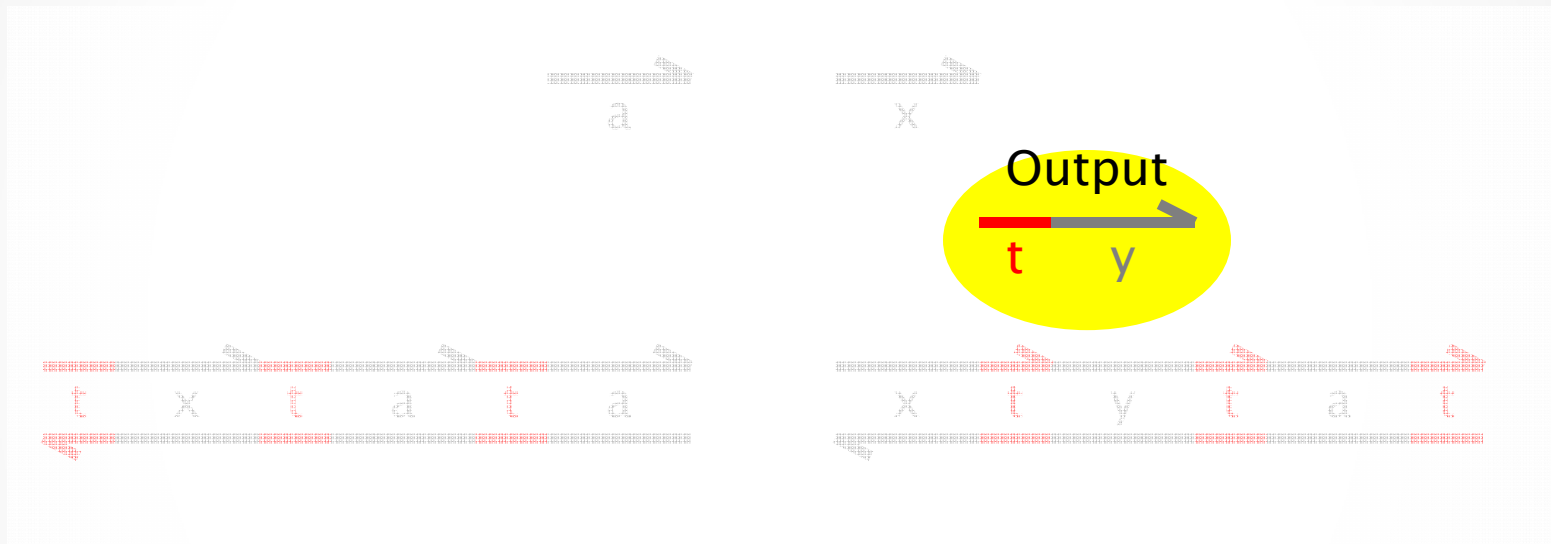




# Transducer $x \rightarrow y$

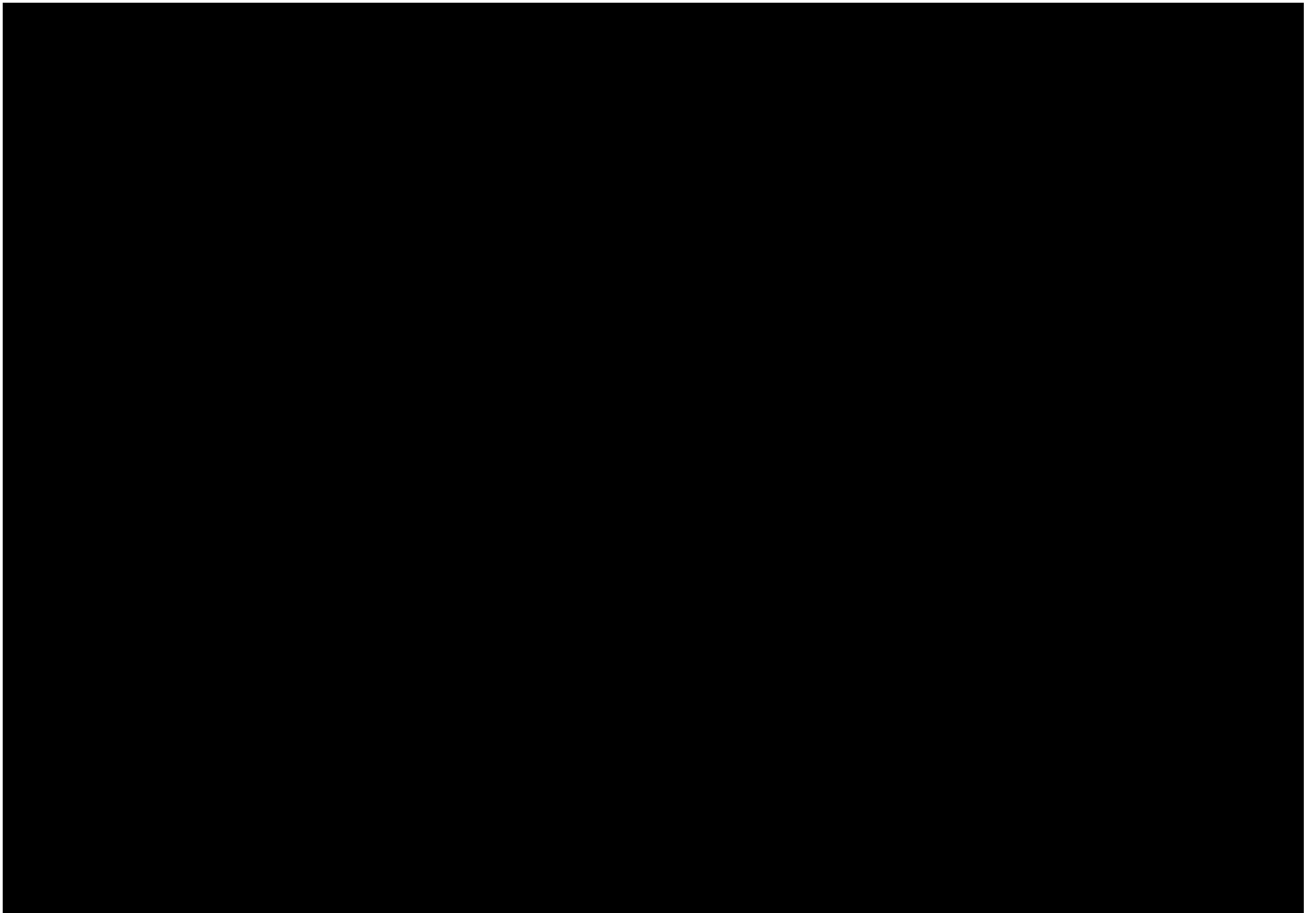


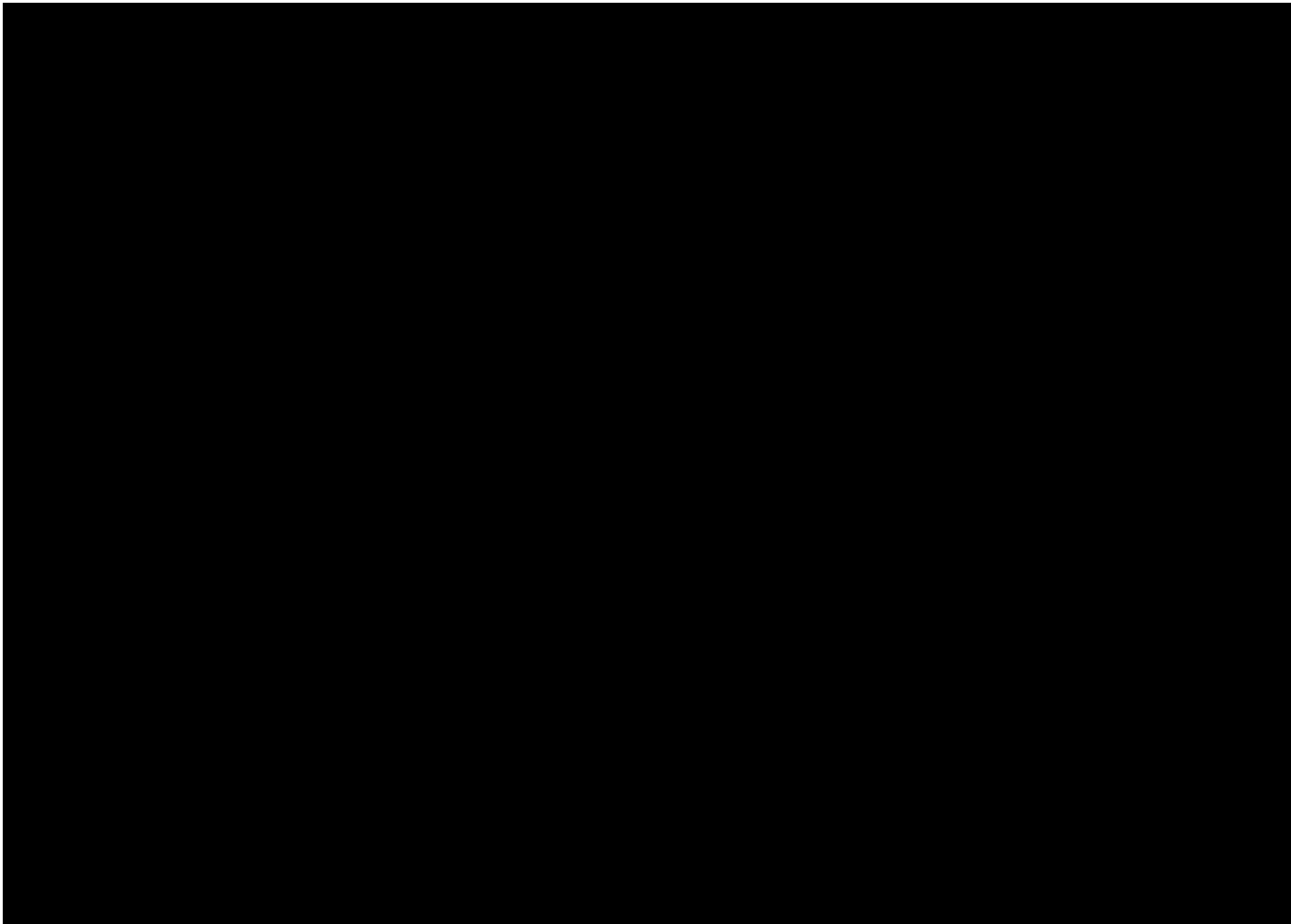
# Transducer $x \rightarrow y$



Done.

N.B. the gate is consumed: it is the energy source.





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# General $n \times m$ Join-Fork

- Easily generalized to 2+ inputs (with 1+ collectors).
- Easily generalized to 2+ outputs.

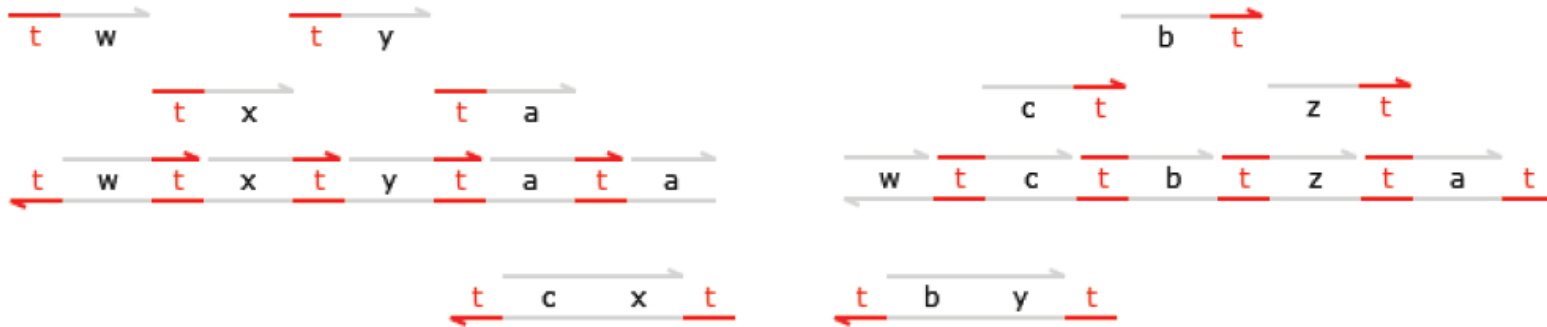


Figure 9: 3-Join  $J_{wxyz} \mid tw \mid tx \mid ty \rightarrow tz$ : initial state plus inputs  $tw, tx, ty$ .

# DNA Programming

Examples:  Compile Simulate Analyse Pause Compilation: Default Options: Simulation: Deterministic View: License Install

Code DNA Input

```
def bind = kt*1.0e-9 (* /nM/s *)
def unbind = kt*exp_DeltaG_over_RT (* /s *)
new t@bind,unbind
new u@bind,unbind
new f1@0.0,0.0

def onex = 50.0

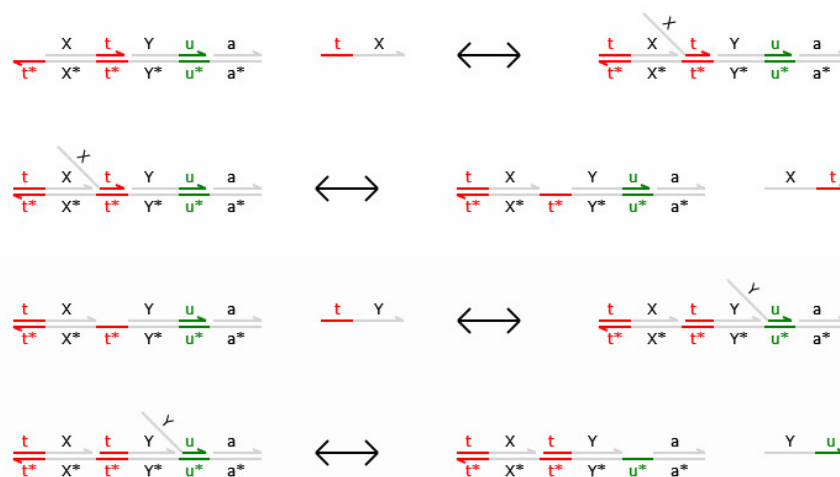
(* x + y -> y + z *)
def Cat(N, x, y, z) =
new a
( (1.5*N) * t^:[x t^]:[y u^]:[a]
| (1.5*N) * [x]:[t^ z]:[t^ y]:u^
| (2.0*N) * <u^ a>
| (2.0*N) * <z t^>
)

def Rep(N,x,f1) =
((3.0*N) * t^:[x]<f1^>)

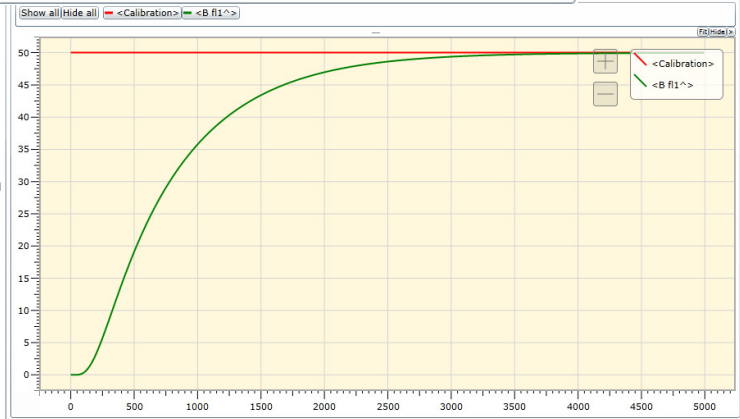
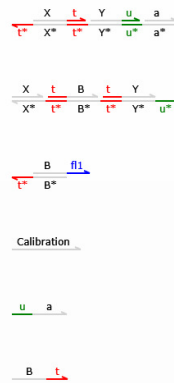
( onex * <Calibration>
| Cat(onex,X,Y,B)
| Rep(onex,B,f11)
| onex * <t^ X>
| onex * <t^ Y>
)
```

Compilation Simulation Analysis

Species Reactions Graph Text Domains SBML



Ready Ln 34 Col 16 Ch 16 INS 100%



# Debugging

- **Big Networks**
  - Two-domain DNA gates for 1 Approximate Majority switch.
  - Initial species: 17
  - Total number of species: 85 (including run-time produced ones)
  - Total number of reactions: 104
- **Analysis**
  - Gate correctness
  - Circuit correctness
  - Compiler correctness
  - Currently, by simulation
  - Increasingly, by modelchecking:

Design and Analysis of DNA  
Strand Displacement Devices  
using Probabilistic Model  
Checking

Matthew R. Lakin <sup>\*†</sup> David Parker <sup>‡†</sup>

Luca Cardelli<sup>\*</sup> Marta Kwiatkowska <sup>‡</sup>

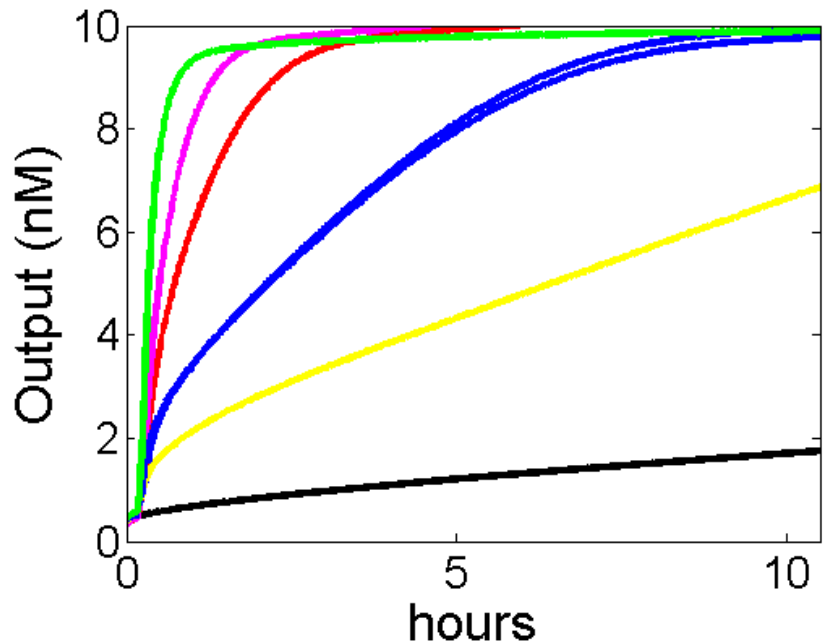
Andrew Phillips<sup>\*§</sup>



# Experiments

Two-domain gate  
for  $X+Y \rightarrow Y+B$

$X+Y \rightarrow Y+B$   
35C  
1x = 50nM



Y  
1x  
0.3x  
0.2x  
0.1x  
0.05x  
0x

Yuan-Jyue Chen and Georg Seelig  
U.Washington.

	$X+Y \rightarrow Y+B$	Concentration
LG1	$\begin{array}{c} X \xrightarrow{T} Y \xrightarrow{U1} a \\ \leftarrow T^* \quad \leftarrow X^* \quad \leftarrow T^* \quad \leftarrow Y^* \quad \leftarrow U1^* \quad \leftarrow a^* \end{array}$	1.5x
LG2	$\begin{array}{c} X \xrightarrow{T} B \xrightarrow{T} Y \\ \leftarrow X^* \quad \leftarrow T^* \quad \leftarrow B^* \quad \leftarrow T^* \quad \leftarrow Y^* \quad \leftarrow U1^* \end{array}$	1.5x
input	$\begin{array}{c} T \xrightarrow{X} \end{array}$	1x
Catalyst	$\begin{array}{c} T \xrightarrow{Y} \end{array}$	0x, 0.05x, 0.1x, 0.2x, 0.3x, 1x
~B	$\begin{array}{c} B \xrightarrow{T} \end{array}$	2x
R1	$\begin{array}{c} U1 \xrightarrow{a} \end{array}$	2x
B readout	$\begin{array}{c} B \xrightarrow{RO} ROX \\ \leftarrow T^* \quad \leftarrow B^* \end{array}$	3x



# Summary

- Executable chemistry

- Given an arbitrary finite chemical network, compile it systematically and execute it.

[D. Soloveichik, G. Seelig, E. Winfree. DNA as a Universal Substrate for Chemical Kinetics. PNAS 107 no. 12, 5393–5398, 2010.]

- Finite chemical networks have the computing power of (stochastic) Petri Nets. Population protocols (such as AM) are also well-characterized. [D. Angluin, J. Aspnes, D. Eisenstat, E. Ruppert: The Computational Power of Population Protocols].

- Executable bio-chemistry

- In addition, DNA supports polymerization, which gives the computing power of Turing Machines.
- Then the programming language cannot be just chemical reactions, but has to be something more like process algebra or term-rewriting systems.

# Conclusions

# Much to be done

- **Systems Biology**

- Develop the algorithmic understanding of molecular networks that will allow us to understand their structure and function (and how to do it better).

- **Synthetic Biology**

- Develop the materials and technology that will allow us to ‘code-up’ arbitrary molecular networks.
- Develop the quantitative techniques that will allow us to ‘debug’ them.

# Acknowledgments

- Microsoft Research
  - Andrew Phillips
- Caltech
  - Winfree Lab
- U.Washington
  - Seelig Lab
- CoSBI
  - Attila Csikász-Nagy



# Challenges

# Verification

- Environment

- The nano-environment is messy (stochastic noise, failures, etc.)
- But we should at least ensure our designs are *logically correct*

- Verifying Components

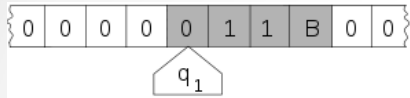
- Reversible reactions (infinite traces)
- Interferences (deadlocks etc.) between copies of the same gate
- Interferences (deadlocks etc.) between copies of different gates
- Removal of active byproducts (garbage collection) is tricky

- Verifying Populations

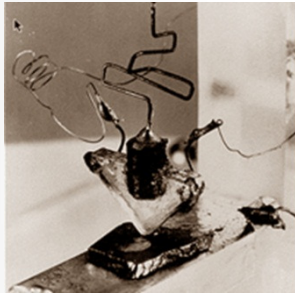
- Gates come in (large) populations
- Each population *shares private domains* (technologically unavoidable)
- Correctness of populations means proofs with large state spaces

# A Brief History of DNA

Turing Machine, 1936



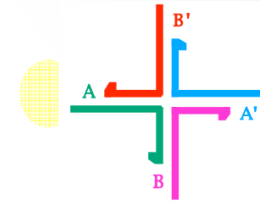
Transistor, 1947



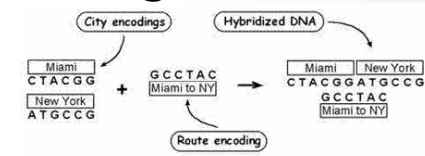
DNA, -3,800,000,000



Structural DNA, 1982



DNA Algorithm, 1994



~~Digital Computers~~  
**Computer programming**

*systematic manipulation of information*  
20<sup>th</sup> century

*systematic manipulation of matter*  
21<sup>th</sup> century

~~DNA Computers~~  
**Molecular programming**



# Conclusions

# Conclusions

- A vast literature on cell cycle switching
  - Ferrell et.al., Novak–Tyson et.al., etc.  
Mostly ODE based analysis, plus noise
  - Many bistable transitions have different implementations in different cell cycle phases and organisms (phosphorylation, enzymes, synthesis/degradation, etc.)
  - We focused on a mechanism that can only be seen stochastically (quick majority switching with  $x=y$ )
- A range of ‘network transformation’
  - Can explain the structure of some natural networks
  - From some non-trivial underlying algorithms
  - Discovering the transformation can elucidate the structure and function of the networks
  - But how can we say that these transformations ‘preserve (essential) behavior’?